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Guidelines for Environmental Infection Control

Recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee

Prepared by

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The material in this report originated in the National Center for Infectious Diseases, James M. Hughes, M.D., Director; and the Healthcare Infection Control Practices Advisory Committee, Robert A. Archer, M.D., Director.

Summary

*The health-care facility environment is rarely implicated in disease transmission, except for inadvertent exposures to environmental pathogens (e.g., *Aspergillus* spp. and *Legionella* spp. for tuberculosis and varicella-zoster virus) can result in adverse patient outcomes and can be prevented by infection-control strategies and engineering controls. Pseudo-outbreaks can be minimized by 1) appropriate use of cleaners and disinfectants; 2) use of automated endoscope reprocessors or hydrotherapy equipment; 3) adherence to water safety standards for specialized care environments (e.g., airborne infection isolation rooms, procedural rooms); 4) management of water intrusion into the facility. Routine environmental sampling is not recommended in hemodialysis settings and other situations where sampling is directed by epidemiologic and control decisions.*

This report reviews previous guidelines and strategies for preventing environmental infection and makes recommendations. These include 1) evidence-based recommendations supported by studies from the Centers for Disease Control and Prevention, U.S. Environmental Protection Agency, U.S. Department of Labor, Occupational Safety and Health Administration, U.S. Department of Justice; 2) guidelines and standards from the American Society of Heating, Refrigerating and Air Conditioning Engineers, American Society of Mechanical Engineers, and American Society of Environmental Engineers; 3) guidelines and standards from building and equipment professional organizations (e.g., International Brotherhood of Electrical Workers, International Association for the Advancement of Medical Instrumentation, and American Society of Environmental Engineers); 4) recommendations derived from scientific theory or rationale; and 5) experienced opinion.

The report also suggests a series of performance measurements as a means of monitoring and improving environmental infection control.

Introduction

Parameters of the Report

This report, which contains the complete list of recommendations with pertinent references, is available on the CDC website. The full four-part guidelines will be available on the CDC website. Relative to previous CDC guidelines:

- revises multiple sections (e.g., cleaning and disinfection of environmental surfaces, regulated medical waste) from previous editions of CDC's *Guideline for Infection Control in Health-Care Facilities*.
- incorporates discussions of air and water environmental concerns from CDC's *Guideline for Infection Control in Health-Care Facilities*.
 - consolidates relevant environmental infection-control measures.
- includes two topics not addressed in previous CDC guidelines --- infection-control measures for water quality in hemodialysis units.

In the full guidelines, Part I, Background Information: Environmental Infection Control, provides a review of the relevant scientific literature. Attention is given to engineering and infection control in the design, renovation, and repair of health-care facilities. Use of an infection-control risk assessment is discussed, as are other activities expected to generate dust or water aerosols. Also reviewed in Part I are catastrophic events (e.g., flooding, sewage spills, loss of electricity and ventilation, and environmental surfaces, laundry, plants, animals, medical wastes, cloth furnishings, and other facilities). Part III and Part IV of the full guidelines provide references (for the full guidelines) for the full guidelines. Part II (this report) contains recommendations for environmental infection control in health-care facilities to prevent infections associated with air, water, or other elements of the environment. This report was developed by a 12-member group that advises CDC on concerns related to the surveillance, prevention, and control of infections primarily in U.S. health-care facilities. In 1999, HICPAC's infection-control focus was primarily on health care is provided (e.g., outpatient surgical centers, urgent care centers, clinics, and nursing facilities). The topics addressed in this report are applicable to the majority of health-care facilities intended for use primarily by infection-control practitioners, epidemiologists, environmental service professionals, information systems professionals, administrators, environmental service professionals, and other health-care workers. The topics addressed in this report are applicable to the majority of health-care facilities intended for use primarily by infection-control practitioners, epidemiologists, environmental service professionals, information systems professionals, administrators, environmental service professionals, and other health-care workers. The topics addressed in this report are applicable to the majority of health-care facilities intended for use primarily by infection-control practitioners, epidemiologists, environmental service professionals, information systems professionals, administrators, environmental service professionals, and other health-care workers.

- infection-control impact of ventilation system and air filtration
- establishment of a multidisciplinary team to conduct infection control risk assessments
- use of dust-control procedures and barriers during construction and renovation
 - environmental infection-control measures for special procedures
- use of airborne-particle sampling to monitor the effectiveness of infection-control procedures to prevent airborne contamination in operating rooms when in use
- guidance regarding appropriate indications for routine culturing of water as part of infection control
 - guidance for recovering from water-system disruptions, water leakage, and other water-related problems
- infection-control concepts for equipment using water from main lines (e.g., water supply equipment, dental unit water lines, and automated dispensing machines)
 - environmental surface cleaning and disinfection strategies with respect to water systems
 - infection-control procedures for health-care workers
 - use of animals in health care for activities
 - managing the presence of service animals in health-care facilities
 - infection-control strategies for when animals receive treatment

• a call to reinstate the practice of inactivating amplified cultures and stocks of
 Topics outside the scope of this report include 1) noninfectious adverse events (e.g., si
 home, 3) home health care, 4) terrorism, and 5) health-care
 Wherever possible, the recommendations in this report are based on data from well-de
 were conducted by using narrowly defined patient populations or specific health-care
 making generalization of findings potentially problematic. Construction standards for
 residential home-care units. Similarly, infection-control measures indicated for imm
 those facilities where such patients are r
 Other recommendations were derived from knowledge gained during infectious disea
 termination of the outbreak was often the result of multiple interventions, the majo
 evaluated. This is especially true for construction situat
 Other recommendations were derived from empiric engineering concepts and may
 conclusions. Where recommendations refer to guidance from the American Institut
 intended for new construction or renovation. Existing structures and engineered syste
 standards in effect at the time of constructio
 Also, in the absence of scientific confirmation, certain infection-control recommend
 strong theoretic rationale and suggestive evidence. Finally, certain recommenda
 Performance Measurements
 Infections caused by the microorganisms described in this guideline are rare events, an
 a facility may not be readily measurable. Therefore, the following steps to mea
 recommendations:
 1. Document whether infection-control personnel are actively involved in all phas
 renovation. Activities should include performing a risk assessment of the neces
 and documenting of the presence of negative airflow within th
 2. Monitor and document daily the negative airflow in AII rooms and positive ai
 rooms.
 3. Perform assays at least once a month by using standard quantitative methods fo
 for heterotrophic and mesophilic bacteria in water used to prepare
 4. Evaluate possible environmental sources (e.g., water, laboratory solutions, or re
 mycobacteria (NTM) of unlikely clinical importance are isolated from clinical cu
 the probable mechanism
 5. Document policies to identify and respond to water damage. Such policies shou
 porous materials within 72 hours, or removal of the wet mater
 Updates to Previous Recommendati
 Contributors to this report reviewed primarily English-language manuscripts identifi
 Medicine's MEDLINE, bibliographies of published articles, and infection-control t
 opinions of all reviewers. This report updates the following publi
 CDC. Guideline for handwashing and hospital environmental control. MMWR 1998;
 laundry, infective waste, and housek
 Tablan OC, Anderson LJ, Arden NH, et al., Hospital Infection Control Practices Adv
 pneumonia. Infect Control Hosp Epidemiol 1994;15:587--627. Updates and expa

aspergillosis and Legionnaires disease; online version incorporates Appendices B, C, and D, and
Legionella spp.

CDC. Guidelines for preventing the transmission of *Mycobacterium tuberculosis* in health care facilities. *MMWR* 2000;49(No. RR12). Supplemental information on engineering controls.

CDC. Recommendations for preventing the spread of vancomycin resistance: recommendations from the National Nosocomial Infection Survey (NNIS) and the Hospital Infection Control Practices Advisory Committee (HICPAC). *MMWR* 1995;44(No. RR12). Supplements environmental controls in hospitals with endemic VRE or continued VRE.

Garner JS, Hospital Infection Control Practices Advisory Committee. Guideline for disinfection and sterilization in Part II --- Recommendations for laundry, routine and terminal cleaning, airborne infection control.

Mangram AJ, Horan TC, Pearson ML, Silver LC, Jarvis WR, Hospital Infection Control Practices Advisory Committee. Guidelines for the prevention of surgical site infection. *Infect Control Hosp Epidemiol* 1999;24:286-300. Cleaning/disinfection recommendations from the section, Intraoperative infection control.

U.S. Public Health Service, Infectious Diseases Society of America, Prevention of Opportunistic Infections in Persons Infected with Human Immunodeficiency Virus. Guidelines for the prevention of opportunistic infections in persons infected with human immunodeficiency virus. *MMWR* 2002; 51:3-64. Supplements information regarding patient interaction.

CDC, Infectious Diseases Society of America, American Society of Blood and Marrow Transplantation. Guidelines for the prevention of opportunistic infections among hematopoietic stem cell transplant recipients. *Cytotherapy* 2001;3:3-12. Infection Control.

Key Terms

Airborne infection isolation (AII) refers to the isolation of patients infected with organisms that are transmitted by the airborne route. This isolation area receives numerous air changes per hour (ACH) (≥ 12 for construction before 2001), and is under negative pressure, such that the direction of airflow is from the corridor into the room. The air in an AII room is preferably exhausted to the outside and filtered through a high-efficiency particulate air (HEPA) filter. The use of personal respiratory protection is required in these rooms when caring for TB or smallpox patients and for staff who lack immunity to varicella-zoster virus [VZV] infection).

Protective environment (PE) is a specialized patient-care area, usually in a hospital, with controlled airflow from the room to the outside adjacent space). The combination of HEPA filtration, negative pressure, and minimal leakage of air into the room creates an environment that can safely accommodate patients with hematopoietic stem cell transplant (HSCT) and immunocompromised patients.

Immunocompromised patients are those patients whose immune mechanisms are diminished due to human immunodeficiency virus [HIV] infection or congenital immune deficiency syndrome (e.g., severe combined immunodeficiency), cardiac failure), or immunosuppressive therapy (e.g., radiation, cytotoxic chemotherapy).

Immunocompromised patients who are identified as high-risk patients have the greatest risk of opportunistic infections. Patients in this subset include persons who are severely neutropenic (ANC of ≤ 500 cells/mL), allogeneic HSCT patients, and those who have received chemotherapy for myelogenous leukemia patient.

Abbreviations

AAMI Association for the Advancement of Medical Instrumentation

ACH air changes per hour

AER automated endoscope repro
AHJ authority having jurisdic
AIA American Institute of Archi
AI airborne infection isolatio
ANSI American National Standards
ASHRAE American Society of Heating, Refrigeration, and
BMBL Biosafety in Microbiological and Biomedical Laboratories
CFR Code of Federal Regulatio
CJD Creutzfeldt-Jakob diseas
CPL compliance document (OSI
DFA direct fluorescence assa
DHHS U.S. Department of Health and Hu
DOT U.S. Department of Transpor
EC environment of care
EPA U. S. Environmental Protection
FDA U.S. Food and Drug Administ
HBV hepatitis B virus
HEPA high efficiency particulate
HIV human immunodeficiency v
HSCT hematopoietic stem cell tran
HVAC heating, ventilation, air cond
ICRA infection-control risk assess
JCAHO Joint Commission on Accreditation of Hea
NaOH sodium hydroxide
NTM nontuberculous mycobact
OSHA Occupational Safety and Health A
PE protective environment
PPE personal protective equipm
TB tuberculosis
USC United States Code
USDA U.S. Department of Agricu
UV ultraviolet
UVGI ultraviolet germicidal irrad
VHF viral hemorrhagic fever
VRE vancomycin-resistant *Enteroco*
VRSA vancomycin-resistant *Staphyloco*
VZV varicella zoster virus

Recommendations for Environmental Infection Control

Rationale for Recommendations

As in previous CDC guidelines, each recommendation is categorized on the basis of expected economic effect. The recommendations are evidence-based wherever possible.

empiric infection-control or engineering principles, theoretic rationale, or from experience (e.g., floods).

The HICPAC system for categorizing recommendations has been modified to include those by state or federal regulations. Guidelines and standards published by the American Society of Heating, Refrigerating and Air Conditioning Engineers (ASHRAE), and the Association for the Advancement of Medical Instrumentation (AIA) are used to make recommendations. These standards reflect a consensus of expert opinions and extensive research. Health and Human Services. Compliance with these standards is usually voluntary. Federal agencies may adopt standards as regulations. For example, the standards from AIA regarding construction have been adopted by reference by >40 states. Certain recommendations have two categories (Evidence-Based and IC), indicating the recommendation is evidence-based as follows:

Rating Categories

Recommendations are rated according to the following categories:

Category IA. Strongly recommended for implementation and strongly supported by scientific studies.

Category IB. Strongly recommended for implementation and supported by certain expert opinion and theoretic rationale.

Category IC. Required by state or federal regulation, or representing an established practice. Agency and regulatory citations are listed where appropriate. Recommendations from AIA guidelines cite the appropriate standard.

Category II. Suggested for implementation and supported by suggestive clinical evidence. Unresolved issue. No recommendation is offered. No consensus or insufficient evidence.

Recommendations --- Air

I. Air-Handling Systems in Health-Care Facilities

A. Use AIA guidelines as minimum standards where state or local regulations are more stringent. Ensure that existing structure and systems in new or renovated health-care facilities. Ensure that existing structure and systems meet minimum standards for new construction (I). Category IC (AIA: 7.2.C7, 9.31.D, 10.31.D, 11.31.D, Environmental Protection Agency).

B. Monitor ventilation systems in accordance with engineers' and manufacturers' recommendations for optimal performance for removal of particulates, and elimination of excess moisture. Category IB, IC (AIA: 7.2.C7, 9.31.D, 10.31.D, 11.31.D, Environmental Protection Agency).

1. Ensure that heating, ventilation, air conditioning (HVAC) filters are properly maintained and replaced on schedule. Category IB, IC (AIA: 7.2.C7, 9.31.D, 10.31.D, 11.31.D, Environmental Protection Agency).

2. Monitor areas with special ventilation requirements (e.g., AII or PE) for AIA standards. Category IB, IC (AIA: 7.2.C7, 9.31.D, 10.31.D, 11.31.D, Environmental Protection Agency).

a. Develop and implement a maintenance schedule for ACH, pressure differential, and humidity control data as part of the multidisciplinary assessment. Take into account the age and condition of the system.

b. Document these parameters, especially the humidity control.

3. Engineer humidity controls into the HVAC system and monitor the control system.

(AIA: 7.31.D9)

- a. Locate duct humidifiers upstream from
 - b. Incorporate a water-removal mechanism
 - c. Locate all duct takeoffs sufficiently downstream from the humidifiers.
 4. Incorporate steam humidifiers, if possible, to reduce potential for microbial growth in ductwork and humidifiers. Category I
 5. Ensure that air intakes and exhaust outlets are located properly in construction zones. Category IC (AIA: 7.31.D1, 7.31.D2, 7.31.D3, 9.31.D3, 10.31.D3, 11.31.D3)
 - a. Locate exhaust outlets >25 ft from air intakes
 - b. Locate outdoor air intakes ≥ 6 ft above ground level
 - c. Locate exhaust outlets from contaminated areas above roof level
 6. Maintain air intakes and inspect filters periodically to ensure proper operation
 7. Bag dust-filled filters immediately upon removal to prevent dispersion of dust. Category IB (AIA: 4.28)
 - a. Seal or close the bag containing the filters
 - b. Discard spent filters as regular solid waste, regardless of the location
 8. Remove bird roosts and nests near air intakes to prevent mites and fungal growth. Category IB
 9. Prevent dust accumulation by cleaning air-duct grilles in accordance with facility cleaning schedule. Grilles are not occupied by patients (AIA: 1.27). Category IC, II (AIA: 7.31.D1, 7.31.D2, 7.31.D3, 7.31.D4, 7.31.D5, 7.31.D6, 7.31.D7, 7.31.D8, 7.31.D9, 7.31.D10, 7.31.D11, 7.31.D12, 7.31.D13, 7.31.D14, 7.31.D15, 7.31.D16, 7.31.D17, 7.31.D18, 7.31.D19, 7.31.D20, 7.31.D21, 7.31.D22, 7.31.D23, 7.31.D24, 7.31.D25, 7.31.D26, 7.31.D27, 7.31.D28, 7.31.D29, 7.31.D30, 7.31.D31, 7.31.D32, 7.31.D33, 7.31.D34, 7.31.D35, 7.31.D36, 7.31.D37, 7.31.D38, 7.31.D39, 7.31.D40, 7.31.D41, 7.31.D42, 7.31.D43, 7.31.D44, 7.31.D45, 7.31.D46, 7.31.D47, 7.31.D48, 7.31.D49, 7.31.D50, 7.31.D51, 7.31.D52, 7.31.D53, 7.31.D54, 7.31.D55, 7.31.D56, 7.31.D57, 7.31.D58, 7.31.D59, 7.31.D60, 7.31.D61, 7.31.D62, 7.31.D63, 7.31.D64, 7.31.D65, 7.31.D66, 7.31.D67, 7.31.D68, 7.31.D69, 7.31.D70, 7.31.D71, 7.31.D72, 7.31.D73, 7.31.D74, 7.31.D75, 7.31.D76, 7.31.D77, 7.31.D78, 7.31.D79, 7.31.D80, 7.31.D81, 7.31.D82, 7.31.D83, 7.31.D84, 7.31.D85, 7.31.D86, 7.31.D87, 7.31.D88, 7.31.D89, 7.31.D90, 7.31.D91, 7.31.D92, 7.31.D93, 7.31.D94, 7.31.D95, 7.31.D96, 7.31.D97, 7.31.D98, 7.31.D99)
 10. Periodically measure output to monitor system function; clean ventilation system to maintain optimum performance (AIA: 1.31, 1.32). Category IC, II (AIA: 7.31.D1, 7.31.D2, 7.31.D3, 7.31.D4, 7.31.D5, 7.31.D6, 7.31.D7, 7.31.D8, 7.31.D9, 7.31.D10, 7.31.D11, 7.31.D12, 7.31.D13, 7.31.D14, 7.31.D15, 7.31.D16, 7.31.D17, 7.31.D18, 7.31.D19, 7.31.D20, 7.31.D21, 7.31.D22, 7.31.D23, 7.31.D24, 7.31.D25, 7.31.D26, 7.31.D27, 7.31.D28, 7.31.D29, 7.31.D30, 7.31.D31, 7.31.D32, 7.31.D33, 7.31.D34, 7.31.D35, 7.31.D36, 7.31.D37, 7.31.D38, 7.31.D39, 7.31.D40, 7.31.D41, 7.31.D42, 7.31.D43, 7.31.D44, 7.31.D45, 7.31.D46, 7.31.D47, 7.31.D48, 7.31.D49, 7.31.D50, 7.31.D51, 7.31.D52, 7.31.D53, 7.31.D54, 7.31.D55, 7.31.D56, 7.31.D57, 7.31.D58, 7.31.D59, 7.31.D60, 7.31.D61, 7.31.D62, 7.31.D63, 7.31.D64, 7.31.D65, 7.31.D66, 7.31.D67, 7.31.D68, 7.31.D69, 7.31.D70, 7.31.D71, 7.31.D72, 7.31.D73, 7.31.D74, 7.31.D75, 7.31.D76, 7.31.D77, 7.31.D78, 7.31.D79, 7.31.D80, 7.31.D81, 7.31.D82, 7.31.D83, 7.31.D84, 7.31.D85, 7.31.D86, 7.31.D87, 7.31.D88, 7.31.D89, 7.31.D90, 7.31.D91, 7.31.D92, 7.31.D93, 7.31.D94, 7.31.D95, 7.31.D96, 7.31.D97, 7.31.D98, 7.31.D99)
- C. Use portable, industrial-grade HEPA filter units capable of filtration rates in excess of 99.97% for respirable particles as needed (AIA: 1.33).
1. Select portable HEPA filters that can recirculate all or nearly all of the room air. Category II
 2. Portable HEPA filter units placed in construction zones can be used later in the project if the filter units are cleaned, and the filter unit performance verified by appropriate parties
 3. Situate portable HEPA units with the advice of facility engineers to ensure

4. Ensure that fresh-air requirements for the area

D. Follow appropriate procedures for use of areas with through-the-wall ventilation (10.31.D18, 11.31.D15)

1. Do not use such areas as PE rooms (1). Category I

2. Do not use a room with a through-the-wall ventilation unit as an AII room unless the engineering controls are met (1,34). Category I (7.2.C3)

E. Conduct an infection-control risk assessment (ICRA) and provide an adequate number of PE rooms to meet the needs of the patient population (1,2,7,8,17,19, 20,34,36).

F. When ultraviolet germicidal irradiation (UVGI) is used as a supplemental engineering control, 1) on the ceiling or suspended from the ceiling as an upper air unit; 2) in the air-return ducts; 3) in PE rooms and booths for sputum induction (34).

G. Seal windows in buildings with centralized HVAC systems, including laboratories (34).

H. Keep emergency doors and exits from PE rooms closed except during an emergency (34). Category II

I. Develop a contingency plan for backup capacity in the event of a general power outage (34). Accreditation of Healthcare Organizations [JCAHO]:

1. Emphasize restoration of appropriate air quality and ventilation conditions in PE rooms, laboratories, departments, and intensive care units (34). Category IC (AIA: 1.5.A1; JCAHO: EC 1.1)

2. Deploy infection-control procedures to protect occupants until power and systems are restored (34). Category IC (AIA: 5.1, 5.2; JCAHO: EC 1.1)

J. Do not shut down HVAC systems in patient-care areas except for maintenance or construction (1,46). Category IB, IC (AIA: 5.1, 5.2)

1. Coordinate HVAC system maintenance with infection-control staff and room closures (34). Category IC (AIA: 5.1, 5.2)

2. Provide backup emergency power and air-handling and pressurization system components (34). Provide pressure differentials in PE rooms, AII rooms, laboratories, and other critical-care areas (1,37,47). Category I (7.2.C3)

3. For areas not served by installed emergency ventilation and backup systems, use portable HEPA-filtered emergency ventilation for patients in those areas (33). Category I (7.2.C3)

4. Coordinate system startups with infection-control staff to protect patients (34). Category IC (AIA: 5.1, 5.2)

5. Allow sufficient time for ACH to clean the air once the system is operational (34). Category IC (AIA: 5.1, 5.2)

- K. HVAC systems serving offices and administrative areas may be shut down for e
alter or adversely affect pressure differentials maintained in laboratories or crit
PE rooms, AII rooms, operating room**
- L. Whenever possible, avoid inactivating or shutting down the entire HVAC s**
- M. Whenever feasible, design and install fixed backup ventilation systems for no
operating rooms, and other critical-care areas identified by**
 - II. Construction, Renovation, Remediation, Rep**
- A. Establish a multidisciplinary team that includes infection-control staff to coordin
consider proactive preventive measures at the inception; produce and mainta**
 - 16,38,48--51). Category IB, IC (**
- B. Educate both the construction team and health-care staff in immunocomprom
risks associated with construction projects, dispersal of fungal spores during su
fungal spores (11--16,27,50,52--56).**
- C. Incorporate mandatory adherence agreements for infection control into const
mechanisms to ensure timely correction of problems (1,11,)**
- D. Establish and maintain surveillance for airborne environmental disease (e.
renovation, repair, and demolition activities to ensure the health and safety of**
 - 1. Using active surveillance, monitor for airborne infections in immuno**
 - 2. Periodically review the facility's microbiologic, histopathologic, and postmorte**
 - IB**
 - 3. If cases of aspergillosis or other health-care--associated airborne fungal infec
biopsies and cultures as feasible**
 - 16,27,50,57--59). Categori**
- E. Implement infection-control measures relevant to construction, renovation, main
IB, IC (AIA: 5.1, 5.2)**
 - 1. Before the project gets under way, perform an ICRA to define the scope of t
16,48--51,60). Category IB, IC (**
 - a. Determine if immunocompromised patients may be at risk for exposure to f
16,48,51).**
 - b. Develop a contingency plan to prevent such**
 - 2. Implement infection-control measures for external demolition and cons**
 - a. Determine if the facility can operate temporarily on recirculate**
 - b. If this is not possible or practical, check the low-efficiency (roughing) filter
of particulates.**
 - c. Seal windows and reduce wherever possible other sources of outside air**

especially in PE areas.

3. Avoid damaging the underground water system (i.e., buried pipes) to prevent
IB, IC (AIA: 5.1)
4. Implement infection-control measures for internal construction activities (
 - a. Construct barriers to prevent dust from construction areas from entering patient-care areas and prevent the spread of fungal spores and in compliance with applicable codes (1,45,48,49,55,64)
 - b. Seal off and block return air vents if rigid barriers are used
 - c. Implement dust-control measures on surfaces and divert pedestrian traffic
 - d. Relocate patients whose rooms are adjacent to work zones, depending on the location of the work zone and the generation of dust or water aerosols and the methods used to control these aerosols
5. Perform those engineering and work-site related infection-control measures for major renovation projects (1,48,49,51,64,66). Categories 5.1, 5.2
 - a. Ensure proper operation of the air-handling system in the affected area after construction is completed and maintain negative pressure (39,47,50,64).
IB
 - b. Create and maintain negative air pressure in work zones adjacent to patient-care areas and ensure that air pressure is maintained (1,48,49,51,64)
 - c. Monitor negative airflow inside rigid barriers
 - d. Monitor barriers and ensure integrity of the construction barriers; 1) monitor for leaks; 2) monitor for damage; 3) repair as needed
 - e. Seal windows in work zones if practical; use window chutes for disposal of debris and maintain negative pressure differential for work zones (1,13,48)
 - f. Direct pedestrian traffic from construction zones away from patient-care areas
 - g. Provide construction crews with 1) designated entrances, corridors, and elevators (with hand hygiene facilities) and convenience services (e.g., vending machines); 2) protective clothing (e.g., coveralls, footwear, and headgear) and a changing anteroom for changing clothing and equipment (1,11,13--16)
 - h. Clean work zones and their entrances daily by 1) wet-wiping tools and toolboxes; 2) cleaning mats with tacky surfaces inside the work zone; and 3) covering debris and securing this covering before removing it
 - i. In patient-care areas, for major repairs that include removal of ceiling tile, use a negative pressure enclosure with plastic sheets or prefabricated plastic enclosure to contain dust; use a negative pressure system within this enclosure to remove dust

portable HEPA filter capable of filtering 300--800 ft³/min., or exhaust air directly to the outdoors.

j. Upon completion of the project, clean the work zone according to facility procedures to remove debris before removing rigid barriers (16,48--50).

k. Flush the water system to clear sediment from pipes to minimize damage.

l. Restore appropriate ACH, humidity, and pressure differential; clean or disinfect surfaces.

F. Use airborne-particle sampling as a tool to evaluate barrier effectiveness.

G. Commission the HVAC system for newly constructed health-care facilities and retrofits on ensuring proper ventilation for operating rooms, AII rooms, and PE areas.

H. No recommendation is offered regarding routine microbiologic air sampling beyond the occupancy of areas housing immunocompromised patients (see 27,48,76,79,80).

I. If a case of health-care--acquired aspergillosis or other opportunistic environmental infection occurs after construction, implement appropriate follow-up measures:

1. Review pressure-differential monitoring documentation to verify that pressure differentials are appropriate for their settings: Category IB, IC (AIA: 27,48,76,79,80).

2. Implement corrective engineering measures to restore proper pressure differentials.

3. Conduct a prospective search for additional cases and intensify retrospective review of laboratory records (27,48,76,79,80).

4. If no epidemiologic evidence of ongoing transmission exists, continue routine surveillance for fungal disease (27,75). Category IB.

J. If no epidemiologic evidence exists of ongoing transmission of fungal disease, continue surveillance for the source (11,13--16,27,44,49--51,60,81,82).

1. Collect environmental samples from potential sources of airborne fungal spores more frequently than settle plates (2,4,11,12,16,27,44,49,50,64,65,81--86). Category IB.

2. If either an environmental source of airborne fungi or an engineering problem is identified, promptly perform corrective measures to eliminate the source and route of entry (49,60). Category IB.

3. Use an EPA-registered antifungal biocide (e.g., copper-8-quinolinolate) for disinfection of surfaces (49,60). Category IB.

4. If an environmental source of airborne fungi is not identified, review infection control procedures to identify potential areas for correction (88,89). Category IB.

5. If possible, perform molecular subtyping of *Aspergillus* spp. isolated from the source to determine identities (90--94). Category IB.

K. If air-supply systems to high-risk areas (e.g., PE rooms) are not optimal, use portable HEPA units until rooms with optimal air-handling systems become available.

III. Infection Control and Ventilation Requirements

A. Minimize exposures of severely immunocompromised patients (e.g., solid-organ transplant recipients) to activities that might cause aerosolization of fungal spores (e.g., vacuuming or dusting).

B. Minimize the length of time that immunocompromised patients in PE are out of their rooms for activities (37,62). Category IC.

C. Provide respiratory protection for severely immunocompromised patients when they are out of their rooms for activities; consult the most recent revision of CDC's *Guideline for Prevention of Nosocomial Pneumonia* regarding the appropriate type of respiratory protection.

D. Incorporate ventilation engineering specifications and dust-controlling procedures (Figure 1). Category IB,

1. Install central or point-of-use HEPA filters for supply (incoming) air (1,2,27,44,100,101). Category IB, IC (Table 7.2.D)

2. Ensure that rooms are well-sealed by 1) properly constructing windows, doors, and doors that are smooth and free of fissures and crevices; 3) sealing walls above and below the ceiling; and 4) monitoring for air leaks (1,27,44,100,101). Category IB, IC (Table 7.2)

3. Ventilate the room to maintain ≥ 12 ACH (1,27,37,100,101). Category IB, IC (Table 7.2)

4. Locate air supply and exhaust grilles so that clean, filtered air enters from one side of the room and exhaust air exits from the opposite side of the room (1,27,100,101). Category IC (Table 7.2)

5. Maintain positive room air pressure (≥ 2.5 Pa [0.01-inch water gauge]) in relation to adjacent areas (Table 7.2)

6. Maintain airflow patterns and monitor these on a daily basis by using permanent airflow indicators in newly constructed or renovated construction, or by using temporary airflow indicators in existing PE units. Document monitoring results.

7. Install self-closing devices on all room exit doors in PE rooms.

E. Do not use laminar air flow systems in newly constructed PE rooms.

F. Take measures to protect immunocompromised patients who would benefit from isolation from other patients with disease (e.g., acute VZV infection or other highly contagious disease).

1. Ensure that the patient's room is designed to receive and return air from an anteroom.

2. Use an anteroom to ensure appropriate air-balance relationships and provide appropriate air filtration. Place a HEPA filter in the exhaust air duct. Return air must be recirculated (1,100) (Figure 2). Category IC.

3. If an anteroom is not available, place the patient in AII and use portable, industrial-grade HEPA filtration in the room (33). Category IC.

G. Maintain backup ventilation equipment (e.g., portable units for fans or filters) in designated areas and take immediate steps to restore the fixed ventilation system if it fails.

IV. Infection-Control and Ventilation Requirements

A. Incorporate certain specifications into the planning and construction or renovation of new or existing AII rooms.

IC

- 1. Maintain continuous negative air pressure (2.5 Pa [0.01 inch water gauge]) in AII rooms. Check negative air pressure periodically, preferably with audible manometers or smoke tubes at the door (for existing AII rooms), or with smoke tubes at the door for new rooms. Document the results of monitoring. (1,100,101). Category IC (AIA: 7.2.1.1).**
- 2. Ensure that rooms are well-sealed by properly constructing windows, doors, and other openings. Use smoke tubes to indicate air leakage, locate the leakage, and make necessary repairs (1,99,100). Category IC (AIA: 7.2.1.2).**
- 3. Install self-closing devices on all AII room exit doors.**
- 4. Provide ventilation to ensure ≥ 12 ACH for renovated rooms and new rooms, including the anteroom. (1,34,105--107). Category IC (AIA: Table 7.2.1).**
- 5. Direct exhaust air to the outside, away from air-intake and populated areas. Do not recirculate air after passing through a HEPA filter. Category IC (AIA: Table 7.2.1).**

B. Where supplemental engineering controls for air cleaning are indicated from a risk assessment, install HEPA filtration in the exhaust air ducts of the HVAC system to supplement HEPA filtration or install HEPA filtration in the room air (34). Category IC (AIA: 7.2.1.3).

C. Implement environmental infection-control measures for persons with diagnosed infectious diseases.

- 1. Use AII rooms for patients with or suspected of having an airborne infectious disease. Enclose the patient in a negative pressure enclosed booth that is engineered to maintain a negative air pressure of ≥ 2.5 Pa (0.01 inch water gauge) with respect to all surrounding spaces with a minimum air change rate of ≥ 50 ft³/min; and 3) air exhausted directly outside away from air intake and recirculation (1,34,105--107). Category IC (AIA: 7.15.E, 7.31.D23, 9.10, 9.11).**
- 2. Although airborne spread of viral hemorrhagic fever (VHF) has not been documented, install an anteroom for a VHF patient in an AII room, plus an anteroom, to reduce the risk of occupational exposure to aerosolized respiratory secretions present in the anteroom during the end stage of a patient's illness (1,34,105--107). Category IC (AIA: 7.15.E, 7.31.D23, 9.10, 9.11).**
 - a. If an anteroom is not available, use portable, industrial-grade HEPA filtration.**

equivalents for removing airborne
b. Ensure that health-care workers wear face shields or goggles with appropriate eye protection, and wear gowns and gloves with prominent cough, vomiting, or hemorrhage ([109](#))

3. Place smallpox patients in negative pressure rooms at the onset of their illness ([36](#)). Category II

D. No recommendation is offered regarding negative pressure or isolation for patients with smallpox. Unresolved issue.

E. Maintain backup ventilation equipment (e.g., portable units for fans or filters) for all rooms, and take immediate steps to restore the fixed ventilation system if it fails.

V. Infection-Control and Ventilation Requirements

A. Implement environmental infection-control and ventilation measures for patients with TB

1. Maintain positive-pressure ventilation with respect to corridors and adjacent rooms.

2. Maintain ≥ 15 ACH, of which ≥ 3 ACH should be fresh air ([127](#)).

3. Filter all recirculated and fresh air through the appropriate filters, providing at least 100% efficiency for particles of 0.3 μ m. Category IC (AIA: Table 7.31.D3)

4. In rooms not engineered for horizontal laminar airflow, introduce air at the ceiling. Category IC (AIA: 7.31.D4)

5. Do not use ultraviolet (UV) lights to prevent surgical-site infections.

6. Keep operating room doors closed except for the passage of equipment, personnel, or patients ([127,128](#)). Category IB

B. Follow precautionary procedures for infectious TB patients who also require anesthesia

1. Use an N95 respirator approved by the National Institute for Occupational Safety and Health (NIOSH) in a room with negative pressure relative to the adjacent rooms ([129,131](#)). Category IC (OC)

Safety and Health Administration [OSHA]; 29 Code of Federal Regulations

2. Intubate the patient in either the AII room or the operating room; if intubating in the operating room, ensure that the operating room doors are closed until 99% of the airborne contaminants are removed ([Table 1](#)) ([34,117](#)).

3. When anesthetizing a patient with confirmed or suspected TB, place a bacterial filter in the patient's breathing circuit to prevent contamination of the operating room airway to prevent contamination of the operating room with equipment or discharge of tubercle bacilli into the air.

4. Extubate and allow the patient to recover in an AII room.

5. If the patient has to be extubated in the operating room, allow adequate time for the patient to recover in the AII room ([Table 1](#)), because extubation is a high-risk procedure ([34,117](#)). Category IC

- C. Use portable, industrial-grade HEPA filters temporarily for supplemental air circulation for patients who require surgery (33,34,117).
 - 1. Position the units appropriately so that all room air passes through the filters at appropriate placements (34). Category I
 - 2. Switch the portable unit off during the surgical procedure. Category I
 - 3. Provide fresh air as per ventilation standards for operating rooms; portable units are not recommended for ACH (1,33,133). Category I
- D. If possible, schedule TB patients as the last surgical cases of the day to maximize Category II
- E. No recommendation is offered for performing orthopedic implant operations. Unresolved issue
- F. Maintain backup ventilation equipment (e.g., portable units for fans or filters) and take immediate steps to restore the fixed ventilation system (1,47)
- VI. Other Potential Infectious Aerosol Hazards in the Operating Room
 - A. In settings where surgical lasers are used, wear appropriate personal protective equipment to minimize exposure to laser plumes (129,135,136). Category I
 - B. Use central wall suction units with in-line filters to evacuate minimal amounts of plume. Category I
 - C. Use a mechanical smoke evacuation system with a high-efficiency filter to manage plume from ablating tissue infected with human papilloma virus (HPV) or performing laser surgery (34,136,137,139--141). Category I
- Recommendations --- Water
 - I. Controlling the Spread of Waterborne Microorganisms
 - A. Practice hand hygiene to prevent the hand transfer of waterborne pathogens, and follow standard precautions (36,142--146). Category I
 - B. Eliminate contaminated water or fluid environmental reservoirs (e.g., in equipment). Category I
 - C. Clean and disinfect sinks and wash basins on a regular basis by using an EPA-registered disinfectant. Category I
 - D. Evaluate for possible environmental sources (e.g., potable water) of specimen contamination if NTM) of unlikely clinical importance are isolated from clinical cultures (e.g., in postprocedural, colonization after use of tap water in patient-care areas). Category I
 - E. Avoid placing decorative fountains and fish tanks in patient-care areas; ensure that fountains are used in public areas of the health-care facility. Category I
 - II. Routine Prevention of Waterborne Microbial Contamination
 - A. Maintain hot water temperature at the return at the highest temperature allowed (≥51°C), and maintain cold water temperature at <68°F (<20°C) (27,147). Category I
 - B. If the hot water temperature can be maintained at ≥124°F (≥51°C), explore engineering solutions (e.g., point-of-use fixtures) to help minimize the risk of Legionnaires' disease. Category I
 - C. When state regulations or codes do not allow hot water temperatures above the minimum 110°F (35°C--43.3°C) for nursing care facilities or when buildings cannot be retrofitted, explore alternative preventive measures to minimize the growth of *Legionella* spp. Category I

1. Periodically increase the hot water temperature to $\geq 150^{\circ}\text{F}$ ($\geq 60^{\circ}\text{C}$)
2. Alternatively, chlorinate the water and then flush it through the system

D. Maintain constant recirculation in hot-water distribution systems serving

III. Remediation Strategies for Distribution System

- A. Whenever possible, disconnect the ice machine before plan
- B. Prepare a contingency plan to estimate water demands for the entire facility expected to result in extensive and heavy microbial or chemical contamination (45,156). Category IC (JCAHO)
- C. When a significant water disruption or an emergency occurs, adhere to any advisory order (157). Category IB, IC (Municipal)

1. Alert patients, families, staff, and visitors not to consume water from drinking fountains while the advisory is in effect, unless the water has been disinfected (e.g., by bringing to a rolling boil for ≥ 1 minute)
2. After the advisory is lifted, run faucets and drinking fountains at full flow for 15 minutes. If chlorination (153,157). Category IC, order; ASHRAE: 12:2000

D. Maintain a high level of surveillance for waterborne disease among patients

E. Corrective decontamination of the hot water system might be necessary after a contamination event has occurred.

1. Decontaminate the system when the fewest occupants are present in the building (ASHRAE: 12:2000)
2. If using high-temperature decontamination, raise the hot-water temperature progressively flushing each outlet until the temperature is $\geq 180^{\circ}\text{F}$ ($\geq 82^{\circ}\text{C}$) for 15 minutes. Category I
3. If using chlorination, add enough chlorine, preferably overnight, to achieve a free chlorine residual of 1 mg/L in the hot water system (153). Category IC (ASHRAE: 12:2000)

a. Flush each outlet until chlorine residual is 1 mg/L

b. Maintain the elevated chlorine concentration in the system for 15 minutes

4. Use a thorough flushing of the water system instead of chlorination if a highly resistant organism (e.g., *Legionella* spp.) is suspected as the water contamination source. Category II

F. Flush and restart equipment and fixtures according to manufacturer's instructions

G. Change the pretreatment filter and disinfect the dialysis water system with an reverse osmosis membrane and downstream microbial

H. Run water softeners through a regeneration cycle to restore t

I. If the facility has a water-holding reservoir or water-storage tank, consult the f whether this equipment needs to be drained, disinfected with an EPA

J. Implement facility procedures to manage a sewage system failure or flooding temporary transfer of patients or provision of services), and establish communic health department to ensure that advisories are received in a timely manner Municipal order)

K. Implement infection-control measures during sewage intrusion, fl

1. Relocate patients and clean or sterilize supplies fr

2. If hands are not visibly soiled or contaminated with proteinaceous material. process 1) before performing invasiv

2) before and after each patient contact; and 3) whenever ha

3. If hands are visibly soiled or contaminated with proteinaceous material, use s

4. If the potable water system is not affected by flooding or sewage contaminatio to standard procedures. Cate

5. Contact the manufacturer of the automated endoscope reprocessor (AER) for water advisory. Category

L. Remediate the facility after sewage intrusion, flooding, o

1. Close off affected areas during cleanup pr

2. Ensure that the sewage system is fully functional before beginning remedia removed. Category II

3. If hard-surfaced equipment, floors, and walls remain in good repair, ensure according to standard cleaning p Category II

4. Clean wood furniture and materials (if still in good repair); allow them to d coatings. Category II

5. Contain dust and debris during remediation and repair as outlined in

M. Regardless of the original source of water damage (e.g., flooding versus water absorbent structural items (e.g., carpeting, wallboard, and wallpaper) and cl cleaned and dried within 72 hours (e.g., moisture content $\leq 20\%$ as determined b soon as the underlying structure is declared by the facility engineer to

IV. Additional Engineering Measures as Indicated by Epidemiologic Investigation Legionnaires Disease

A. When using a pulse or one-time decontamination method, superheat the water b 170°F (71°C--77°C) or hyperchlorinate the system by flushing all outlets for ≥ 5

residual chlorine using a chlorine-based product registered by the EPA for water treatment (153,155,161--164). Category IC

B. After a pulse treatment, maintain both the heated water temperature at the recommendation (Water: II A) wherever practical and permitted by state code (ppm) free residual chlorine at the tap by using a chlorine-based product registered with the EPA (e.g., sodium hypochlorite [bleach]) (153,165--169). Category IC

C. Explore engineering or educational options (e.g., install preset thermostatic mixers at each outlet) to minimize the risk of scalding for patients.

D. No recommendation is offered for treating water in the facility's distribution system with copper or silver, monochloramines, ozone, or UV light (Category IC).

V. General Infection-Control Strategies for Preventing Legionnaires Disease

A. Conduct an infection-control risk assessment of the facility to determine if patients at risk for Legionnaires disease are present (27,189,190). Category IC

B. Implement general strategies for detecting and preventing Legionnaires disease in severely immunocompromised patients (i.e., facilities that do not have HSCT or solid-organ transplant). Category IB

1. Establish a surveillance process to detect health-care--associated Legionnaires disease.

2. Inform health-care personnel (e.g., infection control, physicians, patient-care staff) of the risk of Legionnaires disease to occur and measures to prevent and control health-care--associated legionellosis.

3. Establish mechanisms to provide clinicians with laboratory tests (e.g., culture, PCR, and serology) for the diagnosis of Legionnaires disease (27,189). Category II

C. Maintain a high index of suspicion for health-care--associated Legionnaires disease in suspected cases, especially in patients at risk who do not require antimicrobial therapy: patients aged ≥ 65 years; or patients with chronic underlying disease (e.g., diabetes, chronic lung disease) (27,166,190,192--198).

D. Periodically review the availability and clinicians' use of laboratory diagnostic tests. If the use of the tests on patients with diagnosed or suspected pneumonia is limited, enhance clinicians' use of the test(s) (181,189,191,193,199,200).

E. If one case of laboratory-confirmed, health-care--associated Legionnaires disease or one suspected, health-care-associated Legionnaires disease occur during a 6-month period (181,189,191,193,199,200). Category IC

1. Report the cases to state and local health departments within 24 hours.

2. If the facility does not treat severely immunocompromised patients, conduct a periodic review of microbiologic, serologic, and epidemiologic data to look for previously unidentified cases of health-care--associated Legionnaires disease and surveillance for additional cases.

(27,181,189,191,193,199,200). C

3. If no evidence of continued health-care--associated transmission exists, continue the initiation of surveillance (189,191,193,199,200). Category

F. If there is evidence of continued health-care--associated transmission (i.e., a determine the source of *Legionella* spp. (19

1. Collect water samples from potential aerosolized water sources

2. Save and subtype isolates of *Legionella* spp. obtained from patients and

3. If a source is identified, promptly institute water system decontamination measures

Category IB

4. If *Legionella* spp. are detected in ≥ 1 culture (e.g., conducted at 2-week intervals), decontaminate them accordingly, and repeat decontamination procedures; consider intensive use of techniques used in the and hyperchlorination (27,210,211

IB

G. If an environmental source is not identified during a Legionnaires disease outbreak, Either defer decontamination pending identification of the source of *Legionella* spp. in the water distribution system, with special attention to areas in

H. No recommendation is offered regarding routine culturing of water systems in (i.e., PE or transplant units) for persons at high risk for *Legionella* spp. infection issue

I. No recommendation is offered regarding the removal of faucet aerators in a

J. Keep adequate records of all infection-control measures and environmental

VI. Preventing Legionnaires Disease in Protective Environment

A. When implementing strategies for preventing Legionnaires disease among severely immunocompromised (HSCT or solid-organ transplant) programs, incorporate these specific surveillance strategies outlined previously (see Appendix

1. Maintain a high index of suspicion for legionellosis in transplant patients even if no *Legionella* spp. are cultured (189,215). Category

2. If a case occurs in a severely immunocompromised patient, or if severely immunocompromised cases are identified elsewhere in the facility, conduct a combined epidemiologic investigation of *Legionella* spp. (189,210). Category

B. Implement culture strategies and potable water and fixture treatment measures Category II

1. Depending on state regulations on potable water temperature in public buildings, health-care--associated legionellosis maintain heated water with a minimum return temperature of $\geq 124^{\circ}\text{F}$ ($\geq 51^{\circ}\text{C}$) and add chlorine to water to achieve 1--2 mg/L (1--2 ppm) residual chlorine at the tap (153--155,165,166).
2. Periodic culturing for legionellae in potable water samples from HSCT or solid-organ transplant units as part of a comprehensive strategy to prevent legionellosis in these units (37,154,189,210).
3. No recommendation is offered regarding the optimal methodology (i.e., frequency of sampling, media, and cultures) in HSCT or solid-organ transplant units. Unresolved issue.
4. In areas with patients at risk, when *Legionella* spp. are not detectable in unit water samples, use EPA-registered chlorine disinfectant in aerators monthly by using a chlorine disinfectant (EPA-registered product. If an EPA-registered chlorine disinfectant is not available, use a 1% solution of sodium hypochlorite [v/v dilution]) (153,187). Category IB.

C. If *Legionella* spp. are determined to be present in the water of a transplant unit, the water should be replaced and no longer detected by culture.

1. Decontaminate the water supply as outlined previously (Water Treatment Manual).
2. Do not use water from the faucets in patient-care rooms to avoid cross-contamination.
3. Restrict severely immunocompromised patients from tap water.
4. Use water that is not contaminated with *Legionella* spp. for HSCT.
5. Provide patients with sterile water for tooth brushing, drinking, and for flushing (37,219). Category IB.

D. Do not use large-volume room air humidifiers that create aerosols (e.g., by Venturi effect) in patient-care rooms. If used, they should be subjected to high-level disinfection and filled only with sterile water.

VII. Cooling Towers and Evaporative Condensers

- A. When planning construction of new health-care facilities, locate cooling towers away from patient-care areas, and design the towers to minimize the volume of aerosol drift.
- B. Implement infection-control procedures for operational cooling towers.

1. Install drift eliminators (153,203,222). Category IB.
2. Use an effective EPA-registered biocide on a regular basis.
3. Maintain towers according to manufacturers' recommendations, and keep records of maintenance, including environmental test results from towers. Investigate towers during outbreak investigations (153). Category IB.

C. If cooling towers or evaporative condensers are implicated in health-care--associated legionellosis, they should be replaced (199,203,221,223). Category IB.

VIII. Dialysis Water Quality and Disinfection

- A. Adhere to current AAMI standards for quality-assurance performance of devices in hemodialysis centers (both acute and maintenance [chronic] settings) and for Category IA, IC (AAMI: American National Standards Institute [ANSI/AAMI] RD47:1993, RD62:2001).**
- B. No recommendation is offered regarding whether more stringent requirements for hemodiafiltration. Unresolved.**
- C. Conduct microbiologic testing specific to water in dialysis settings (229,230,236).**
 - 1. Perform bacteriologic assays of water and dialysis fluids at least once a month using standard methods (236--238). Category IA, IC (AAMI: ANSI/AAMI RD62:2001).**
 - a. Assay for heterotrophic, mesophilic bacteria using membrane filtration methods.**
 - b. Do not use nutrient-rich media (e.g., blood agar).**
 - 2. In conjunction with microbiologic testing, perform endotoxin testing on product water (229,230,239--242). Category IA, IC (AAMI: ANSI/AAMI RD5:1992, ANSI/AAMI RD47:1993).**
 - 3. Ensure that water does not exceed the limits for microbial counts and endotoxin (AAMI: ANSI/AAMI RD5:1992, ANSI/AAMI RD47:1993).**
- D. Disinfect water distribution systems in dialysis settings at least weekly (226--228, RD62:2001).**
- E. Wherever practical, design and engineer water systems in dialysis settings to avoid dead-end branches and taps that can harbor bacteria (226--228,231,236). Category IA, IC.**
- F. When storage tanks are used in dialysis systems, they should be routinely drained and disinfected with an ultrafilter or pyrogenic filter (membrane filter with a pore size sufficient to remove bacteria) in the water line distal to the storage tank (236). Category IA, IC.**

IX. Ice Machines and Ice

- A. Do not handle ice directly by hand, and wash hands before and after handling ice.**
- B. Use a smooth-surface ice scoop to dispense ice.**
 - 1. Keep the ice scoop on a chain short enough that the scoop cannot touch the floor (243,244). Category IA, IC.**
 - 2. Do not store the ice scoop in the ice storage bin.**
- C. Do not store pharmaceuticals or medical solutions on ice intended for consumption.**
- D. Machines that dispense ice are preferred to those that require ice to be removed from equipment specifically manufactured for this purpose.**

- E. Limit access to ice-storage chests, and keep container doors closed
- F. Clean, disinfect, and maintain ice-storage chests on

- 1. Follow the manufacturer's instructions for
- 2. Use an EPA-registered disinfectant suitable for use on ice machines, dispenser
Category II
- 3. If instructions and EPA-registered disinfectants suitable for use on ice mach
regimen ([Box 3](#)) (244). Categ
- 4. Flush and clean ice machines and dispensers if they have not been disconnect
II

- G. Install proper air gaps where the condensate lines me
- H. Conduct microbiologic sampling of ice, ice chests, and ice-making machines a
investigation (244,248,249). Cat
- X. Hydrotherapy Tanks and Po
- A. Drain and clean hydrotherapy equipment (e.g., Hubbard tanks, tubs, whirlpoo
use, and disinfect equipment surfaces and components by using an EPA-regi
instructions. Category I
- B. In the absence of an EPA-registered product for water treatmen

- 1. Maintain a 15-ppm chlorine residual in the water of small hydrotherapy
- 2. Maintain a 2--5-ppm chlorine residual in the water of whirlp
- 3. If the pH of the municipal water is in the basic range (e.g., when chloramine
community), consult the facility engi
the possible need to adjust the pH of the water to a more acidic level before d
(252). Category II

- C. Clean and disinfect hydrotherapy equipment after
- D. Clean and disinfect inflatable tubs unless they are sin
- E. No recommendation is offered regarding the use of antiseptic chemicals (e.g., c
Unresolved issue
- F. Conduct a risk assessment of patients before their use of large hydrotherapy
incontinence from pool use until their conditi
- G. For large hydrotherapy pools, use pH and chlorine residual levels appropriate
agencies. Category IC (Sta
- H. No recommendation is offered regarding the use in health-care settings of w
recreational use. Unresolved

- XI. Miscellaneous Medical Equipment Connecte
- A. Clean, disinfect, and maintain AER equipment according to the manufacturer'
inadvertent contamination of endoscopes and bronchoscopes with wate

1. To rinse disinfected endoscopes and bronchoscopes, use water of the highest quality (e.g., sterile water or bacteriologic water [water filtered through 0.1--0.2- μ m filters])
 2. Dry the internal channels of the reprocessed endoscope or bronchoscope by forced-air treatment) to lessen the proliferation of waterborne microorganisms and to help prevent
- B. Use water that meets nationally recognized standards set by the EPA for drinking water (264--267). Category IC**
- C. Take precautions to prevent waterborne contamination of dental units**

1. After each patient, discharge water and air for a minimum of 20--30 seconds from the dental unit system that enters a patient's mouth through handpieces, ultrasonic scalers, or air/water syringes
 2. Consult with dental water-line manufacturers to 1) determine suitable methods for monitoring the water to ensure quality is maintained; and 2) determine appropriate monitoring methods
 3. Consult with the dental unit manufacturer regarding the need for periodic disinfection
- Category IB**

Recommendations ---Environmental

I. Cleaning and Disinfecting Strategies for Environmental Surfaces

- A. Select EPA-registered disinfectants, if available, and use them in accordance with the manufacturer's instructions (EPA: 7 United States Code [USC])**
- B. Do not use high-level disinfectants/liquid chemical sterilants for disinfection of environmental surfaces; such use is counter to label instructions for these products (FDA: 21 CFR 80)**
- C. Follow manufacturers' instructions for cleaning and maintaining environmental surfaces**
- D. In the absence of a manufacturer's cleaning instructions, use the following strategies:**

1. Clean noncritical medical equipment surfaces with a detergent/disinfectant or a registered hospital disinfectant with a tuberculocidal claim (depending on the nature of the surface and the degree of contamination) (274). Category IC
2. Do not use alcohol to disinfect large environmental surfaces
3. Use barrier protective coverings as appropriate for noncritical surfaces that are 1) likely to be contaminated with blood or body substances; 2) likely to be contaminated with blood or body substances; or 3) difficult to clean

- E. Keep housekeeping surfaces (e.g., floors, walls, tabletops) visibly clean on a regular basis**
1. Use a one-step process and an EPA-registered hospital detergent/disinfectant

care areas where 1) uncertainty exists about the nature of the soil on the surfaces (e.g., blood or body fluid contamination versus the presence of multidrug resistant organisms) on such surfaces (272,274,280,281).

2. Detergent and water are adequate for cleaning surfaces in nonpatient-care areas.
3. Clean and disinfect high-touch surfaces (e.g., doorknobs, bed rails, light switches) on a more frequent schedule than for low-touch housekeeping surfaces. (272,274,280,281).
4. Clean walls, blinds, and window curtains in patient-care areas when they are soiled.

F. Do not perform disinfectant fogging in patient-care areas.

G. Avoid large-surface cleaning methods that produce mists or aerosols, or disperse dust.

H. Follow proper procedures for effective uses of mops, cloths, and brushes.

1. Prepare cleaning solutions daily or as needed, and replace with fresh solution when the solution becomes soiled (280,281). Category II
2. Change the mop head at the beginning of each day and also as required by facility policy for soiled mop heads or other body substances. Category II
3. Clean mops and cloths after use and allow to dry before reuse; or use single-use mops and cloths. Category II

I. After the last surgical procedure of the day or night, wet vacuum or mop operating rooms with a registered hospital disinfectant (114,115).

J. Do not use mats with tacky surfaces at the entrances to operating rooms.

K. Use appropriate dusting methods for patient-care areas designated for immunocompromised patients. Category IB

1. Wet-dust horizontal surfaces daily by moistening a cloth with a small amount of water (37,40,280). Category II
2. Avoid dusting methods that disperse dust (e.g., feathering). Category II

L. Keep vacuums in good repair and equip vacuums with HEPA filters for use in immunocompromised patients' rooms.

M. Close the doors of immunocompromised patients' rooms when vacuuming, wet dusting, or mopping to reduce airborne dust (37,40,289). Category II

N. When performing low- or intermediate-level disinfection of environmental surfaces, avoid direct exposure of neonates to disinfectant residues on these surfaces by using EPA-registered disinfectants and following the manufacturer's instructions and safety advisories (271,290--292). Category II

1. Do not use phenolics or any other chemical germicide to disinfect bassinets. Category IB
2. Rinse disinfectant-treated surfaces, especially those treated with phenolics, with water before use.

O. When using phenolic disinfectants in neonatal units, prepare solutions to conform with manufacturer's instructions, or use premixed formulations (271,290--292). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

II. Cleaning Spills of Blood and Body Fluids

A. Promptly clean and decontaminate spills of blood or other potentially infectious materials (293,298,299,301,302). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

B. Follow proper procedures for site decontamination of spills of blood or blood-contaminated surfaces (293,298,299,301,302). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

1. Use protective gloves and other PPE appropriate for this task (293). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

2. If the spill contains large amounts of blood or body fluids, clean the visible material with a cloth or paper towels moderately wetted with disinfectant solution (293,298,299,301,302). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

3. Swab the area with a cloth or paper towels moderately wetted with disinfectant solution (293,298,299,301,302). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

C. Use germicides registered by the EPA for use as hospital disinfectants and lab disinfectants (293,301,303). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

D. An EPA-registered sodium hypochlorite product is preferred, but if such product is not available, use a household chlorine bleach product (e.g., household chlorine bleach) not containing additives (293,301,303). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

1. Use a 1:100 dilution (500--615 ppm available chlorine) to decontaminate non-porous surfaces (293,301,303). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

2. If a spill involves large amounts of blood or body fluids, or if a blood or culture spill (293,301,303). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

III. Carpeting and Cloth Furnishings

A. Vacuum carpeting in public areas of health-care facilities and in general patient-care areas (44,304,307). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

B. Periodically perform a thorough, deep cleaning of carpeting as determined by manufacturer's instructions (44,304,307). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

C. Avoid use of carpeting in high-traffic zones in patient-care areas or where spills occur (44,304,307). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

D. Follow appropriate procedures for managing spills (44,304,307). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

1. Spot-clean blood or body substance spills promptly (293,301,304,307). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

2. If a spill occurs on carpet tiles, replace any tiles contaminated by blood and body fluids (293,301,304,307). Category IC (OSHA: 29 CFR 1910.1030 § d.4.ii.A)

- E. Thoroughly dry wet carpeting to prevent the growth of fungi; replace carpeting
 - F. No recommendation is offered regarding the routine use of fungicidal or bactericidal cleaning agents in patient-care facility or in general patient-care areas
 - G. Do not use carpeting in hallways and patient rooms in areas housing immunocompromised patients
 - H. Avoid using upholstered furniture and furnishings in high-risk patient-care areas to reduce the risk of contamination (e.g., pediatrics units) (318,320--322)
 - I. No recommendation is offered regarding whether upholstered furniture and furnishings should be cleaned with a process appropriate for the type of upholstery
- Unresolved issue

- 1. Maintain upholstered furniture in good condition
 - 2. Maintain the surface integrity of the upholstery by repairing tears and fraying
 - 3. If upholstered furniture in a patient's room requires cleaning to remove visible soil, clean the furniture in a maintenance area where it can be fully cleaned with a process appropriate for the type of upholstery
- IV. Flowers and Plants in Patient-Care Areas**
- A. Flowers and potted plants need not be restricted from areas for immunocompromised patients
 - B. Designate care and maintenance of flowers and potted plants to staff not directly involved in patient care
 - C. If plant or flower care by patient-care staff is unavoidable, instruct the staff to wear gloves and perform hand hygiene after glove removal (318,320--322)
 - D. Do not allow fresh or dried flowers, or potted plants, in patient-care areas for immunocompromised patients

- V. Pest Control**
- A. Develop pest-control strategies, with emphasis on kitchens, cafeterias, laundry, construction areas, docks, construction activities, and other areas prone to infestation
 - B. Install screens on all windows that open to the outside; keep screens in good condition
 - C. Contract for routine pest control service by a credentialed pest-control specialist (318,320--322). Category IB
 - D. Place laboratory specimens (e.g., fixed sputum smears) in covered containers

- VI. Special Pathogens**
- A. Use appropriate hand hygiene, PPE (e.g., gloves), and isolation precautions during care of patients with special pathogens (318,320--322). Category IB
 - B. Use standard cleaning and disinfection protocols to control environmental contamination by special pathogens (e.g., methicillin-resistant *Staphylococcus aureus*, vancomycin intermediate sensitive *Staphylococcus aureus*, *Enterococcus* [VRE]) (318,320--322). Category IB
- 1. Pay close attention to cleaning and disinfection of high-touch surfaces in patient rooms, including commodes, bed rails, doorknobs, or light switches (318,320--322). Category IB
 - 2. Ensure compliance by housekeeping staff with cleaning and disinfection protocols for high-touch surfaces (318,320--322). Category IB
 - 3. Use EPA-registered chemical germicides appropriate for the surface to be disinfected, as specified by the manufacturer (318,320--322). Category IB

instructions (271,322--327). Category IB, IC (

4. When contact precautions are indicated for patient care, use disposable patient care items to minimize cross-contamination and to reduce the risk of exposure to multiple-resistant microorganisms (3

5. Follow these same surface-cleaning and disinfecting measures for managing the

II

C. Environmental-surface culturing can be used to verify the efficacy of hospital disinfecting rooms that house patients with VRE

1. Obtain prior approval from infection-control staff and the clinical laboratory Category II

2. Infection-control staff, with clinical laboratory staff consultation, must

D. Thoroughly clean and disinfect environmental and medical equipment surfaces in accordance with manufacturers' instructions (271,274,319,334). (

E. Advise families, visitors, and patients regarding the importance of hand hygiene (e.g., respiratory secretions or fecal matter) to s

F. Do not use high-level disinfectants (i.e., liquid chemical sterilants) on environmental surfaces in accordance with manufacturers' instructions because of the toxicity of the chemicals (270,273,274,278

G. Because no EPA-registered products are specific for inactivating *Clostridium difficile*, disinfection of environmental surfaces in accordance with guidance from the surveillance and epidemiology indicate ongoing transmission (

H. No recommendation is offered regarding the use of specific EPA-registered hospital disinfectants for *C. difficile*. Unresolved is:

I. Apply standard cleaning and disinfection procedures to control environmental surfaces in pediatric-care units and care areas for immunocompromised patients (

J. Clean surfaces that have been contaminated with body substances; perform local decontamination with an EPA-registered disinfectant in accordance with the manufacturer's instructions (271,274,319,334); 1910.1030 § d.4.ii.A; EPA: 7 USC §

K. Use disposable barrier coverings as appropriate to minimize contamination of surfaces; 1910.1030 § d.4.ii.A; EPA: 7 USC §

L. Develop and maintain cleaning and disinfection procedures in patient-care areas for Creutzfeldt-Jakob disease (CJD), for which no EPA-registered disinfectants are available (271,274,319,334); 1910.1030 § d.4.ii.A; EPA: 7 USC §

1. In the absence of contamination with central nervous system tissue, extraordinary cleaning and disinfection or applying full-strength sodium hypochlorite are not needed for routine cleaning or terminal disinfection of a room housing a patient with CJD Category II

Category II

2. After removing gross tissue from the surface, use either 1N NaOH or a sodium hypochlorite solution containing 20,000 ppm available chlorine (d

1:5 to 1:3 v/v, respectively, of U.S. household chlorine bleach; contact the man

products for advice) to decontaminate operating room or autopsy surfaces with central nervous system or cerebral spinal fluid (CJD patient (273,337--342). Ca

- a. The contact time for the chemical used during this process
- b. Blot up the chemical with absorbent material and rinse the surface
- c. Discard the used, absorbent material into appropriate waste container

3. Use disposable, impervious covers to minimize body substance contamination

M. Use standard procedures for containment, cleaning, and decontamination (Environmental Services: II) (293). Category IC (OSHA: 29 CFR 1910.1030 § d.3.v)

1. Wear PPE appropriate for a surface decontamination and cleaning task (293)
2. Discard used PPE by using routine disposal procedures or decontaminate reusable PPE (29 CFR 1910.1030 § d.3.v)

Recommendations ---Environmental Services

I. General Information

A. Do not conduct random, undirected, microbiologic sampling of air, water, and surfaces (Environmental Services: I) (293). Category IB

B. When indicated, conduct microbiologic sampling as part of an epidemiologic investigation to determine environmental conditions to detect contamination or verify abatement (Environmental Services: I) (293). Category IB

C. Limit microbiologic sampling for quality assurance purposes to 1) biologic monitoring of air, water, and dialysate in hemodialysis units; and 3) short-term evaluation of the implementation of control protocols (270,343). Category IB

II. Air, Water, and Environmental Surfaces

A. When conducting any form of environmental sampling, identify existing company procedures and use standard methods (343--347). Category IB

B. Select a high-volume air sampling device if anticipated levels of microbiologic contamination are high (345,346,348,349). Category IB

C. Do not use settle plates to quantify the concentration of airborne microorganisms (343,347). Category IB

D. When sampling water, choose growth media and incubation conditions that are appropriate for the microorganisms being sampled (343,347). Category IB

E. When using a sample/rinse method for sampling an environmental surface, develop a protocol for sample collection, transport, and analysis (343,347). Category IB

F. When environmental samples and patient specimens are available for comparison, identify and compare the microorganisms down to the species level at a minimum, and beyond if necessary (343,347). Category IB

Recommendations ---Laundry and Linens

I. Employer Responsibilities

A. Employers must launder workers' personal protective garments or uniforms contaminated with infectious materials (293). Category IC (OSHA: 29 CFR 1910.1030 § d.3.v)

II. Laundry Facilities and Equipment

- A. Maintain the receiving area for contaminated textiles at negative pressure consistent with AIA construction standards in effect during the time of facility construction
- B. Ensure that laundry areas have handwashing facilities and products and appropriate signage (AIA: 7.23.D4; OSHA: 29 CFR 1910.1030 § d.4.iv)
- C. Use and maintain laundry equipment according to manufacturer's instructions
- D. Do not leave damp textiles or fabrics in machines or hampers
- E. Disinfection of washing and drying machines in residential care is not needed and proper washing and drying procedures should be followed

III. Routine Handling of Contaminated Textiles

- A. Handle contaminated textiles and fabrics with minimum agitation to avoid contamination (357--361). Category IC (OSHA: 29 CFR 1910.1030 § d.4.iv)
- B. Bag or otherwise contain contaminated textiles and fabrics at the point of use
 - 1. Do not sort or prerinse contaminated textiles or fabrics in patient-care areas
 - 2. Use leak-resistant containment for textiles and fabrics contaminated with blood or body fluids (357--361). Category IC (OSHA: 29 CFR 1910.1030 § d.4.iv)
 - 3. Identify bags or containers for contaminated textiles with labels, color coding, and other means to identify contents (357--361). Category IC (OSHA: 29 CFR 1910.1030 § d.4.iv)
- C. Covers are not needed on contaminated textile hampers
- D. If laundry chutes are used, ensure that they are properly designed, maintained, and covered to prevent leakage of contaminated laundry (357--361). Category IC (OSHA: 29 CFR 1910.1030 § d.4.iv)
 - 1. Ensure that laundry bags are closed before tossing the fabric into the chute
 - 2. Do not place loose items in the laundry
- E. Establish a facility policy to determine when textiles or fabrics should be sorted by type of material (362,363). Category II

IV. Laundry Process

- A. If hot-water laundry cycles are used, wash with detergent in water $\geq 160^{\circ}\text{F}$ ($\geq 71^{\circ}\text{C}$) for at least 10 minutes (364--365). Category II
- B. No recommendation is offered regarding a hot-water temperature setting and cycle time for laundry in long-term care facilities. Unresolved issue
- C. Follow fabric-care instructions and special laundering requirements
- D. Choose chemicals suitable for low-temperature washing at proper use concentrations (365--370). Category II
- E. Package, transport, and store clean textiles and fabrics by methods that will ensure they remain clean during interfacility loading, transport, and unloading

V. Microbiologic Sampling of Textiles

- A. Do not conduct routine microbiologic sampling of clean textiles

B. Use microbiologic sampling during outbreak investigations if epidemiologic evidence suggests a role for the facility in disease transmission (371). Category I

VI. Special Laundry Situation

- A. Use sterilized textiles, surgical drapes, and gowns for situations requiring sterilization**
- B. Use hygienically clean textiles (i.e., laundered, but not sterilized) in non-sterile situations**
- C. Follow manufacturers' recommendations for cleaning fabric products, including heat-labile items**
- D. Do not use dry cleaning for routine laundering in health-care facilities**
- E. Use caution when considering use of antimicrobial mattresses, textiles, and clothing items; EPA has not approved public health claims asserting protection against hospital-acquired infections**
- F. No recommendation is offered regarding using disposable fabrics and linens**

VII. Mattresses and Pillows

- A. Keep mattresses dry; discard them if they remain wet or stained, particularly if they are used in the intensive care unit**
- B. Clean and disinfect mattress covers by using EPA-registered disinfectants that are compatible with the cover material. Avoid the development of tears, cracks, or holes in the cover**
- C. Maintain the integrity of mattress and pillow covers**

- 1. Replace mattress and pillow covers if they become torn or cracked**
- 2. Do not stick needles into a mattress through the cover**

- D. Clean and disinfect moisture-resistant mattress covers between patient use by using EPA-registered disinfectants that are compatible with the cover material**
- E. If using a mattress cover completely made of fabric, change these covers and pillow covers between patients**
- F. Launder pillow covers and washable pillows in the hot-water cycle between patients**

substances (382). Category I

VIII. Air-Fluidized Beds

- A. Follow manufacturers' instructions for air-fluidized bed maintenance**
- B. Change the polyester filter sheet at least weekly or as indicated by the manufacturer**
- C. Clean and disinfect the polyester filter sheet thoroughly, especially between patients**

Category IB

- D. Consult the facility engineer to determine the proper placement of air-fluidized beds**

Recommendations --- Animals in Health-Care Facilities

I. General Infection-Control Measures for Animals

- A. Minimize contact with animal saliva, dander, urine, and feces**
- B. Practice hand hygiene after any animal contact**

- 1. Wash hands with soap and water, especially if hands are visibly soiled or contaminated**
- 2. Use either soap and water or alcohol-based hand rubs when hands are not visibly soiled**

II. Animal-Assisted Activities and Resident Activities

- A. Avoid selection of nonhuman primates and reptiles in animal-assisted activities (391--393). Category II**
- B. Enroll animals that are fully vaccinated for zoonotic diseases and that are healthy or otherwise have completed recent anthelmintic treatment under the supervision of a licensed veterinarian**

- C. Enroll animals that are trained with the assistance or under the direction of personnel
- D. Ensure that animals are controlled by persons trained in providing activities or and behavior traits (391,394). C
- E. Take prompt action when an incident of biting or scratching by an animal

- 1. Remove the animal permanently from these premises
- 2. Report the incident promptly to appropriate authorities (e.g., infection-control personnel) (391). Category II
- 3. Promptly clean and treat scratches, bites, or other wounds

- F. Perform an ICRA and work actively with the animal handler before conducting the session whether the session should be held in a public area of the facility or in a private area

- G. Take precautions to mitigate allergic responses

- 1. Minimize shedding of animal dander by bathing animals <2 times per week
- 2. Groom animals to remove loose hair before a visit, or use a grooming bag

- H. Use routine cleaning protocols for housekeeping surfaces

- I. Restrict resident animals, including fish in tanks, from access to patient-care areas, sterile supply areas, sterile and clean supply storage areas, medication preparation areas, and other areas

Category II

- J. Establish a facility policy for regular cleaning of fish tanks, rodent cages, and bird cages; assign a cleaning task to a nonpatient-care staff member; avoid splashing tank water on patients and bedding. Category II

III. Protective Measures for Immunocompromised Patients

- A. Advise patients to avoid contact with animal feces, saliva, urine, or other animal secretions
- B. Promptly clean and treat scratches, bites, or other wounds
- C. Advise patients to avoid direct or indirect contact with animals
- D. Conduct a case-by-case assessment to determine if animal-assisted activities or services are appropriate for immunocompromised patients (394)
- E. No recommendation is offered regarding permitting pet visits to terminally ill patients

Unresolved issue.

IV. Service Animals

- A. Avoid providing facility access to nonhuman primates and reptile species
- B. Allow service animals access to the facility in accordance with the Americans with Disabilities Act unless the animal creates a direct threat to other persons or a fundamental alteration of the facility's services. Department of Justice: 28 CFR 36.208
- C. When a decision must be made regarding a service animal's access to any part of the facility, animal, patient, and health-care situation on a case-by-case basis to determine if reasonable modifications in policies and procedures will mitigate this risk (394)

D. If a patient must be separated from his or her service animal while in the health-care facility, arrangements have been made for supervision or care of the animal during the patient's absence and arrangements to address the patient's needs in the absence of the animal.

V. Animals as Patients in Human Health-Care Facilities

A. Develop health-care facility policies to address the treatment of animals in health-care facilities.

- 1. Use the multidisciplinary team approach to policy development, including policy review and updates, and activities. Category II**
- 2. Exhaust all veterinary facility, equipment, and instrument options**
- 3. Ensure that the care of the animal is supervised by a licensed veterinarian.**

B. When animals are treated in human health-care facilities, avoid treating animals with invasive procedures are performed (e.g., cardiac catheterization laboratory).

C. Schedule the animal procedure for the last procedure of the day in the area, at a time when the facility is in the vicinity. Category II

D. Adhere strictly to standard precautions.

E. Clean and disinfect environmental surfaces thoroughly by using an EPA-registered disinfectant. Category II

F. Allow sufficient ACH to clean the air and help remove airborne dander, mold, and dust.

G. Clean and disinfect using EPA-registered products or sterilize equipment that has been used on animals. Category I

H. If reusable medical or surgical instruments are used in an animal procedure, clean and disinfect them. Category II

VI. Research Animals in Health-Care Facilities

A. Use animals obtained from quality stock, or quarantine incoming animals.

B. Treat sick animals or remove them from the facility.

C. Provide prophylactic vaccinations, as available, to animal handlers.

D. Ensure proper ventilation through appropriate facility design and location (399 USC 2131)

- 1. Keep animal rooms at negative pressure relative to corridors**
- 2. Prevent air in animal rooms from recirculating elsewhere in the health-care facility.**

E. Keep doors to animal research rooms closed.

F. Restrict access to animal facilities to essential personnel.

G. Establish employee occupational health programs specific to the animal research procedures specific to zoonoses with occupational health clinics in the health-care facility. Health and Human Services [DHHS]: Biosafety in Microbiological and Biomedical Laboratories (42 CFR 139)

H. Document standard operating procedures for the unit (42 CFR 139)

I. Conduct routine employee training on worker safety concerns relevant to the animal research facility.

animal handling) (400,401). Category IC (DHHS: BMBL;

J. Use precautions to prevent development of animal-induced asthma in an
Recommendations --- Regulated Medic

I. Categories of Regulated Medical

A. Designate the following as major categories of medical waste that require special handling: 1) laboratory wastes [e.g., cultures and stocks of microorganisms]; 2) bulk blood, body fluids, pathology and anatomy waste; and 4) sharps [e.g., needles, scalpels, suture needles]

B. Consult federal, state, and local regulations to determine if other waste items are regulated.
Category IC (States; OSHA: 29 CFR 1910.1030 § g.2.1; Department of Transportation: 49 CFR 173.133 CO23.8)

II. Disposal Plan for Regulated Medical Waste

A. Develop a plan for the collection, handling, predisposal treatment, and terminal disposal of regulated medical waste.
IC (States; OSHA: 29 CFR 1910.1030 § g.2.1)

B. Designate a person or persons as responsible for establishing, monitoring, and evaluating the disposal plan.
III. Handling, Transporting, and Storing Regulated Medical Waste

A. Inform personnel involved in handling and disposal of potentially infective wastes that they and their employees are trained in appropriate handling and disposal methods (293). Category IC (States; OSHA: 29 CFR 1910.1030 § g.2.1)

B. Manage the handling and disposal of regulated medical wastes generated in isolation rooms and other patient-care areas so that they do not become a source of infection.
medical wastes from other patient-care areas are not to be placed in sharps containers.

C. Use proper sharps disposal strategies (293). Category IC (States; OSHA: 29 CFR 1910.1030 § g.2.1)

1. Use a sharps container capable of maintaining its impermeability after waste is placed in it and until final disposal (293). Category IC (States; OSHA: 29 CFR 1910.1030 § d.4.iii.A)

2. Place disposable syringes with needles, including sterile sharps that are being used, in puncture-resistant containers located as close as is practical to the point of use (293). Category IC (States; OSHA: 29 CFR 1910.1030 § d.2.vii)

3. Do not bend, recap, or break used syringe needles before discarding them into a sharps container (293).
1910.1030 § d.2.vii and § d.2.viii

D. Store regulated medical wastes awaiting treatment in a properly ventilated area to prevent development of noxious odors. Category IC (States; OSHA: 29 CFR 1910.1030 § g.2.1)

E. If treatment options are not available at the site where the medical waste is generated, use impervious containers to transport the waste to the on-site treatment location or to another facility for treatment.
IV. Treatment and Disposal of Regulated Medical Waste

A. Treat regulated medical wastes by using a method (e.g., steam sterilization, autoclaving, incineration, or other technology) approved by the appropriate authority having jurisdiction (Agency for Toxic Substances and Hazardous Waste Administration) before disposal in a sanitary landfill or other final disposal site.
Category IC (States; OSHA: 29 CFR 1910.1030 § g.2.1)

B. Follow precautions for treating microbiologic wastes (e.g., amplified cultures and other infectious agents) in a BMBL facility.
BMBL)

1. Biosafety level 4 laboratories must inactivate microbiologic wastes in the lab (autoclaving) before transport to and disposal in a sanitary landfill (400). Category IC
 2. Biosafety level 3 laboratories must inactivate microbiologic wastes in the lab (autoclaving) or incinerate them at a sanitary landfill before transport to and disposal in a sanitary landfill (400).
- C. Biosafety levels 1 and 2 laboratories should develop strategies to inactivate an approved inactivation method (e.g., autoclaving) instead of packaging and shipping and disposal (400,406--408). Category IC
- D. Laboratories that isolate select agents from clinical specimens must comply with the requirements for the isolation, storage, and appropriate disposal of these agents (409). Category I
- E. Sanitary sewers may be used for safe disposal of blood, suctioned fluids, ground water, and other fluids if the following conditions are met: sewage discharge requirements are met and that the state has declared this to be an appropriate disposal method.
- V. Special Precautions for Wastes Generated During Care of Patients with Creutzfeldt-Jakob Disease
- A. When discarding items contaminated with blood and body fluids from VHF patients, use appropriate precautions to prevent splashing and agitation during handling (36,109).
 - B. Manage properly contained wastes from areas providing care to VHF patients in designated areas (Regulated Medical Waste: III B) (36,109).
 - C. Decontaminate bulk blood and body fluids from VHF patients by using approved methods (e.g., autoclaving) before disposal (36,109). Category IC
 - D. When discarding regulated medical waste generated during the routine (i.e., not for diagnostic or research purposes) care of patients (e.g., blood, sharps, or pathological waste) (36,270,271), use appropriate precautions (e.g., autoclaving) before disposal (36,109).
 - E. Incinerate medical wastes (e.g., central nervous system tissues or contaminated instruments) from patients with confirmed or suspected CJD using the standard procedures of diagnosed or suspected CJD patients (36,270,271).

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Figure 1

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Box 1

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